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Exploring QSAR of melatonin receptor ligand benzofuran derivatives using E-state index

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Abstract—Considering the recent challenge to the medicinal chemists for the development of selective melatonin receptor ligands, an attempt has been made to explore physicochemical requirements of benzofuran derivatives for binding with human MT_1 and MT_2 receptor subtypes and also to explore selectivity requirements. In this study, E-states of different common atoms of the molecules (calculated according to Kier and Hall) and physicochemical parameters (partition coefficient and molar refractivity) were used as independent variables along with suitable dummy parameters. The best equation describing MT_1 binding affinity [n = 34, $Q^2 = 0.670$, $R_a^2 = 0.790$, $R^2 = 0.822$, R = 0.907, s = 0.609, F = 25.8 (df = 5, 28)] suggests that the binding affinity decreases as the value of n (number of CH₂ spacer beside R₂) increases while it increases with rise in electrotopological state values of different atoms of the benzofuran ring. Again, presence of methoxy group at R₁ and hydrogen, unsubstituted phenyl or fluoro-substituted phenyl group at R₂ is conducive to the MT_1 binding affinity. The binding affinity decreases if furyl substitution at R₃ position is present. The best equation describing MT_2 binding affinity [n = 34, $Q^2 = 0.602$, $R_a^2 = 0.755$, $R^2 = 0.792$, R = 0.890, s = 0.584, F = 21.3 (df = 5, 28)] shows that the MT_2 binding affinity depends on the similar factors as described for MT_1 binding affinity; however, the contributions of the factors for the two affinities are different to some extent as evidenced from the regression coefficients. Among the selectivity relations, the best equation [n = 33, $Q^2 = 0.496$, $R_a^2 = 0.681$, $R^2 = 0.721$, R = 0.849, s = 0.458, F = 18.1 (df = 4, 28)] suggests that MT_2 binding increases with increase in value of n, presence of methoxy group at R₁, and E-state values of different atoms of the benzofuran ring, while it decreases in presence of furyl group at R₃ position. © 2004 Elsevier Ltd. All rights rese

Melatonin (N-acetyl-5-methoxyindolamine) is a derivative of tryptophan, an essential amino acid for mammals. Earlier reports indicated that melatonin was uniquely synthesized and secreted by the pineal gland in vertebrates, but recent studies¹ revealed its extrapineal origin including retina, gut ovary, testes, bone marrow and lens.

Melatonin has several important physiological actions including the control of circadian rhythms, sleep induction, regulation of seasonal reproduction and immune enhancement.² Melatonin was also shown to have significantly broader actions including oncostatic effects, immune system stimulation and anti-inflammatory functions.³ Melatonin has been shown to possess both

in vitro and in vivo important antioxidant activities. Further, it inhibits the activation of poly (ADP ribose) synthetase, ^{4,5} and several aspects of the inflammatory response. ⁶ Melatonin can reduce the toxicity of a wide variety of environmental and chemical insults, which initiate oxidative stress.

Melatonin is the only known chronobiotic, hormonal regulator of neoplastic cell growth. Melatonin acts as a differentiating agent in some cancer cells and lowers their invasive and metastatic status through alterations in adhesion molecules and maintenance of gap junctional intercellular communication. In some cancer types, melatonin, either alone or in combination with other agents, induces apoptotic cell death.⁷

The MT₁, MT₂ and MT₃ melatonin receptors are possible mediators of the physiological effects of melatonin,⁸ and they have been expressed at central and peripheral sites. Central effects appear to be predominantly mediated by the MT₁ subtype. Activation of the

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MT₁ melatonin receptor inhibits neuronal firing rate in the suprachiasmatic nucleus (SCN) and prolactin secretion from the pars tuberalis and induces vasoconstriction. Activation of the MT₂ melatonin receptor phase shifts circadian rhythms generated within the SCN, inhibits dopamine release in the retina, induces vasodialation, enhances splenocyte proliferation and inhibits leukocyte rolling in the microvasculature. Activation of the MT₃ melatonin receptor reduces intraocular pressure and inhibits leukotriene B4-induced leukocyte adhesion.⁹

Quantitative structure–activity relationship (QSAR) studies have been done on various derivatives acting on melatonin receptors. Comparative molecular field analysis (CoMFA) models have been reported on binding affinity within different classes of melatonin receptors of indole-based melatonin analogs, 10 2-iodomelatonin 11 and N-substituted melatonin derivatives. 12 Topographical model has been used for the QSAR study of 2-N-acylaminoalkylindoles. 13 Partial least squares (PLS) and multiple regression analysis (MRA) were used as tools for QSAR studies on 2-substituted indole compounds. 14

The present paper attempts QSAR modeling of MT₁ and MT₂ receptor binding and explores selectivity requirements for MT₂ versus MT₁ binding of benzofuran derivatives¹⁵ using electrotopological state (E-state) index, developed by Kier and Hall. ^{16,17}

The E-state index is an atom level descriptor 16,17 encoding both electronic character and topological environment of each skeletal atom in a molecule. It is derived from graph theoretic approach and has two basic components: (a) intrinsic topological and electronic state of an atom; (b) effect of the environment influencing the atom (perturbation), considering differences in the intrinsic electrotopological states of different atoms and topological distance among them which determine the magnitude of the interactions. The E-state index has been projected as a useful tool in the context of QSAR studies and reported to have power to identify atoms or fragments in the molecules that are important for the biological activity.¹⁷ The present group of authors also have used E-state index to explore QSARs of ligands acting on different pharmacologically relevant targets of contemporary interest. 18-22 In continuation of such efforts, the present communication will show here the utility of E-state index in QSAR studies by exploring selectivity requirements of MT₂ versus MT₁ binding of benzofuran derivatives using E-state index.

The structural features of the compounds and their melatonin receptor binding affinities are given in Table 1. In the present study, we have tried to incorporate hydrophobicity parameter (partition coefficient $\log P$), steric factor (molar refractivity MR, ignoring its polarizability component) and some indicator or integer variables along with E-state parameters to improve the quality of final relations. The physicochemical parameter values ($\log P$ and MR) were

calculated by CHEM DRAW ULTRA 5.0 software²³ using Crippen's fragmentation method.²⁴ All compounds considered in the present study contain fourteen common atoms (excluding hydrogens). The atoms of the molecules were numbered keeping serial numbers of the common atoms same in all the compounds (as shown in Fig. 1). The E-state index values were calculated using electro1 program.25 The program AUTOREG²⁵ was used to relate binding affinity with different combinations of E-state values of different atoms along with appropriate physicochemical variables and indicator variables in order to find out the best multiple regression relations. For this purpose, all possible combinations of predictor variables were tried with a restriction that predictor variables used in an equation are not much intercorrelated (|r| < 0.5). Using the program RRR98,²⁵ regression coefficients with corresponding standard errors and various statistical parameters reflecting quality²⁶ (like explained variance R_a^2 or adjusted R^2 , correlation coefficient R, standard error of estimate s, variance ratio F and average of absolute values of the residuals AVRES) of the equations were found out. Leave-one-out cross-validation²⁷ was done using the programs KRPRES1 and KRPRES2, 25 which generate predicted variance (Q^2) , predicted residual sum of squares (PRESS), standard deviation of error of prediction (SDEP) and average of absolute values of predicted residuals (Pres_{av}). While deriving the final relations, log P and MR values could not find place in the best equations. Though many relations were generated, only the final ones will be reported here for brevity. The regression coefficients and variance ratios of all reported equations are significant at 95% and 99% levels, respectively. For convenience, definitions of different variables appearing in the reported equations are given in Table 2.

In case of MT₁ binding activity, the best relation involving all 34 compounds was the following:

$$\begin{split} pC_1 &= 3.357(\pm 2.334)S_3 - 1.379(\pm 0.419)n \\ &+ 1.712(\pm 0.591)I_{\text{OMe}} + 1.170(\pm 0.466)I_{\text{Ph/F}} \\ &- 2.342(\pm 0.943)I_{\text{Furyl}} - 5.146 \\ n &= 34, \ \ Q^2 = 0.670, \ \ R_a = 0.790, \ \ R^2 = 0.822, \\ R &= 0.907, \ \ F = 25.8(5,28), \ \ s = 0.609, \\ \text{AVRES} &= 0.433, \ \ \text{SDEP} = 0.751, \\ S_{\text{PRESS}} &= 0.828, \ \ \text{PRESS} = 19.2, \ \ \text{Pres}_{\text{av}} = 0.557 \end{split}$$

The 95% confidence intervals of the regression coefficients are shown within parentheses. Eq. 1 could predict 67.0% and explain 79.0% of the variance of MT_1 binding affinity. As E-state values of atom numbers 1, 2, 3, 4 and 9 are highly intercorrelated (|r| > 0.9), a new variable S_{12349} was defined as sum of E-state values of the atoms. The statistical quality of the resultant equation was comparable to that of Eq. 1.

Table 1. Structural features and melatonin receptor binding affinity of benzofuran derivatives

Sl.		Structural features				Melatonin receptor binding affinity								
No.					MT_1 binding affinity (pC_1)			MT_2 binding affinity (pC_2)			Selectivity (pC_2-pC_1)			
	n	\mathbf{R}_1	R_2	R ₃	Obs.a	Calc.b	Pred.b	Obs.a	Calc.c	Pred.c	Obs.a	Calc.d	Pred.d	
1	0	OCH ₃	Н	CH ₃	3.824	3.990	4.013	3.469	4.129	4.222	-0.355	-0.074	-0.023	
2	1	OCH_3	C_6H_5	CH ₃	2.896	2.724	2.711	4.301	3.745	3.702	1.405	1.546	1.555	
3	1	OCH_3	$C_6H_4(2\text{-OCH}_3)$	CH_3	1.845	1.510	1.480	3.469	3.034	2.994	1.624	1.472	1.463	
4	1	OCH_3	$C_6H_4(3-OCH_3)$	CH_3	1.391	1.518	1.530	3.495	3.053	3.012	2.104	1.489	1.451	
5	1	OCH_3	$C_6H_4(4\text{-}OCH_3)$	CH_3	1.620	1.524	1.516	3.301	3.067	3.045	1.681	1.500	1.489	
6	1	OCH_3	$C_6H_4(3-F)$	CH_3	3.125	2.537	2.483	3.638	3.350	3.320	0.513	1.331	1.383	
7	1	OCH_3	$C_6H_4(3-CF_3)$	CH_3	1.097	1.094	1.094	2.932	2.179	2.033	1.835	1.035	0.878	
8	1	OCH_3	$C_6H_4(3-C1)$	CH_3	2.143	1.498	1.440	3.658	3.010	2.950	1.515	1.474	1.471	
9	1	OCH_3	$C_6H_3(2,6-Cl_2)$	CH_3	1.073	1.414	1.445	1.893	2.829	2.912	0.820	1.359	1.391	
10	1	OCH_3	$C_6H_3(3,5-CF_3)$	CH_3	0.322	0.634	0.766	0.469	1.229	2.085	0.147	0.525	1.223	
11	1	OCH_3	C_6H_{11}	CH_3	1.178	1.689	1.743	2.604	3.427	3.550	1.426	1.712	1.751	
12	1	OCH_3	$C_5H_4N(3)$	CH_3	1.538	1.513	1.511	2.452	3.040	3.095	0.914	1.497	1.533	
13	3	OCH_3	C_6H_5	CH_3	1.502	0.577	-1.053	2.818	3.060	3.459	1.316	_	_	
14	1	C_2H_5	C_6H_5	CH_3	1.191	1.758	1.907	2.084	2.475	2.558	0.893	1.037	1.064	
15	1	C_2H_5	$C_6H_4(3\text{-OCH}_3)$	CH_3	0.541	0.552	0.556	1.770	1.783	1.787	1.229	0.981	0.938	
16	1	Н	C_6H_5	CH_3	0.810	1.572	1.728	1.975	2.295	2.356	1.165	1.024	0.998	
17	1	Н	$C_6H_4(3\text{-OCH}_3)$	CH_3	0.640	0.365	0.280	1.730	1.603	1.566	1.090	0.967	0.946	
18	0	OCH_3	C_6H_5	CH_3	5.000	4.065	3.928	4.699	4.323	4.269	-0.301	-0.069	-0.026	
19	1	OCH_3	C_6H_5	CH_2I	2.062	2.738	2.791	3.387	3.774	3.805	1.325	1.554	1.571	
20	1	OCH_3	C_6H_5	C_3H_7	2.483	2.755	2.776	3.699	3.810	3.818	1.216	1.568	1.594	
21	1	OCH_3	C_6H_5	Furyl	0.156	0.352	0.571	1.435	0.650	-0.225	1.279	0.723	0.039	
22	1	OCH_3	C_6H_5	$CH=CH_2$	1.270	2.697	2.809	3.161	3.688	3.729	1.891	1.503	1.478	
23	1	OCH_3	$C_6H_4(3\text{-OCH}_3)$	$CH=CH_2$	1.260	1.490	1.511	2.559	2.996	3.036	1.299	1.446	1.455	
24	0	OCH_3	C_6H_5	$CH=CH_2$	4.699	4.037	3.942	5.000	4.266	4.162	0.301	-0.112	-0.187	
25	0	OCH_3	C_6H_5	Furyl	1.889	1.693	1.474	0.442	1.227	2.102	-1.447	-0.891	-0.207	
26	1	OCH_3	C_6H_5	CH ₂ CH=CH ₂	2.455	2.718	2.738	4.222	3.733	3.695	1.767	1.524	1.508	
27	1	OCH_3	$C_6H_4(3\text{-OCH}_3)$	$CH_2CH=CH_2$	1.666	1.512	1.498	3.959	3.041	2.955	2.293	1.468	1.419	
28	0	OCH_3	C_6H_5	$CH_2CH=CH_2$	5.000	4.058	3.922	4.699	4.311	4.255	-0.301	-0.090	-0.051	
29	0	OCH_3	C_6H_5	CH=CHCH ₃	2.996	4.065	4.221	3.678	4.323	4.417	0.682	-0.085	-0.225	
30	1	OCH_3	C_6H_5	NHCH ₃	2.270	2.708	2.742	3.456	3.710	3.730	1.186	1.514	1.536	
31	0	OCH_3	C_6H_5	NHCH ₃	4.398	4.048	3.998	4.398	4.288	4.272	0.000	-0.100	-0.118	
32	1	ОН	C_6H_5	CH ₃	0.245	0.080	-0.226	1.097	0.860	0.694	0.852	0.525	0.418	
33	1	OC_5H_{11}	C_6H_5	CH ₃	1.370	1.265	1.246	2.012	2.055	2.063	0.642	0.868	0.906	
34	1	OC_6H_{13}	C_6H_5	CH_3	2.086	1.291	1.146	2.493	2.090	2.017	0.407	0.876	0.955	

^a Obs. = Observed (Ref. 15), Calc. = Calculated, Pred. = Predicted.

$$\begin{split} pC_1 &= 0.584(\pm 0.412)S_{12349} - 1.395(\pm 0.423)n \\ &+ 1.704(\pm 0.592)I_{\rm OMe} + 1.156(\pm 0.469)I_{\rm Ph/F} \\ &- 2.322(\pm 0.946)I_{\rm Furyl} - 3.231 \\ n &= 34, \ \ Q^2 = 0.673, \ \ R_a^2 = 0.789, \ \ R^2 = 0.821, \\ R &= 0.906, \ \ F = 25.6(5,28), \ \ s = 0.611, \\ {\rm AVRES} &= 0.433, \ \ {\rm SDEP} = 0.749, \\ S_{\rm PRESS} &= 0.825, \ \ {\rm PRESS} = 19.1, \ \ {\rm Pres}_{\rm av} = 0.551 \end{split} \label{eq:pressure}$$

The predictor variables used in Eqs. 1 and 2 are not much intercorrelated (Table 3). The variable S_{12349} in Eq. 2 implies the importance of electron density distribution

Figure 1. General structure of benzofuran derivatives: the common atoms have been numbered 1 through 14.

of different atoms of the benzofuran nucleus, which is in turn dependent on the R_1 substituent The negative coefficient of n indicates the MT_1 binding affinity

^bFrom Eq. 1.

^c From Eq. 3.

^d From Eq. 6.

Table 2. Definitions of variables

Variable	Definition
S_3	E-state value of atom no. 3
S_4	E-state value of atom no. 4
S_8	E-state value of atom no. 8
S_{12349}	Sum of E-state values of atom nos. 1, 2, 3, 4 and 9
S_{4589}	Sum of E-state values of atom nos. 4, 5, 8 and 9
$I_{ m OMe}$	Indicator variable having value 1 in presence of
	methoxy group at R_1 position, value 0 otherwise.
$I_{ m Ph/F}$	Indicator variable having value 1 in presence of
	hydrogen, unsubstituted phenyl or fluoro-substituted
	phenyl group at R_2 position, value 0 otherwise.
$I_{ m Furyl}$	Indicator variable having value 1 in presence of furyl
-	substitution at R ₃ position, value 0 otherwise.
n	Number of CH ₂ spacer beside R ₂

decreases on increase of number of CH_2 spacer (n) beside R_2 . Again, the positive coefficients of I_{OMe} and $I_{Ph/F}$ indicate that presence of methoxy group at R_1 and hydrogen, unsubstituted phenyl or fluoro-substituted phenyl group at R_2 , respectively, are conducive to the MT_1 binding affinity. Further, the negative coefficient of I_{Furyl} denotes that MT_1 binding affinity decreases if furyl substitution at R_3 position is present. The calculated and predicted MT_1 binding affinity values according to Eq. 1 are given in Table 1.

In case of MT₂ binding activity, the best relation was the following:

$$pC_2 = 5.742(\pm 2.781)S_4 - 0.670(\pm 0.400)n$$

$$+ 1.988(\pm 0.556)I_{OMe} + 0.615(\pm 0.454)I_{Ph/F}$$

$$- 3.033(\pm 0.904)I_{Furyl} - 9.287$$

$$n = 34, \ Q^2 = 0.602, \ R_a^2 = 0.755, \ R^2 = 0.792,$$

$$R = 0.890, \ F = 21.3(5,28), \ s = 0.584,$$

$$AVRES = 0.466, \ SDEP = 0.733,$$

$$S_{PRESS} = 0.808, \ PRESS = 18.3, \ Pres_{av} = 0.606$$
 (3)

Eq. 3 could predict 60.2% and explain 75.5% of the variance of MT_2 binding affinity. The predictor variables of Eq. 3 are not much intercorrelated (Table 3). As E-state values of atom numbers 4, 5, 8 and 9 are highly intercorrelated (|r| > 0.9), a new variable S_{4589} was defined as sum of E-state values of the atoms. The statistical quality of the resultant equation was comparable to that of Eq. 3.

$$\begin{split} pC_2 &= 1.198(\pm 0.599)S_{4589} - 0.671(\pm 0.406)n \\ &+ 1.924(\pm 0.554)I_{\rm OMe} + 0.564(\pm 0.467)I_{\rm Ph/F} \\ &- 2.955(\pm 0.915)I_{\rm Furyl} - 4.008 \\ n &= 34, \ \ Q^2 = 0.590, \ \ R_{\rm a}^2 = 0.749, \ \ R^2 = 0.787, \\ R &= 0.877, \ \ F = 20.7(5,28), \ \ s = 0.591, \\ {\rm AVRES} &= 0.469, \ \ {\rm SDEP} = 0.744, \\ S_{\rm PRESS} &= 0.820, \ \ {\rm PRESS} = 18.8, \ \ {\rm Pres}_{\rm av} = 0.613 \end{split}$$

The variable S_{4589} in Eq. 4 implies the importance of electron density distribution of the benzofuran nucleus, which is in turn dependent on the R₁ substituent. The impact of the variables n, $I_{\rm OMe}$, $I_{\rm Ph/F}$ and $I_{\rm Furyl}$ shows similar trend as found in case of MT₁ binding but only differing in the numerical value of the corresponding coefficient. This is also supported by the high intercorrelation between MT₂ and MT₁ binding affinity for the present set of compounds (Table 3). The calculated and predicted MT₂ binding affinity values according to Eq. 3 are given in Table 1.

While exploring selectivity relations, compound 13 had to be deleted because of its outlier behavior. The following best relation was obtained:

$$\begin{split} pC_2 - pC_1 &= 1.924(\pm 1.542)S_8 + 1.533(\pm 0.419)n \\ &\quad + 0.716(\pm 0.443)I_{\text{OMe}} - 0.743(\pm 0.697)I_{\text{Furyl}} \\ &\quad - 2.874 \\ n &= 33, \ \ Q^2 = 0.492, \ \ R_a^2 = 0.682, \\ R^2 &= 0.722, \ \ R = 0.850, \ \ F = 18.2(4,28), \\ s &= 0.457, \ \ \text{AVRES} = 0.357, \ \ \text{SDEP} = 0.569, \\ S_{\text{PRESS}} &= 0.618, \ \ \text{PRESS} = 10.7, \ \ \text{Pres}_{\text{av}} = 0.460 \end{split}$$

As E-state values of atom numbers 4, 5, 8, 9 are highly intercorrelated (|r| > 0.9), S_{4589} as defined previously was used:

$$pC_2 - pC_1 = 0.575(\pm 0.465)S_{4589} + 1.551(\pm 0.420)n$$

$$+ 0.744(\pm 0.453)I_{OMe} - 0.754(\pm 0.698)I_{Furyl}$$

$$- 3.612$$

$$n = 33, Q^2 = 0.492, R_a^2 = 0.681,$$

$$R^2 = 0.721, R = 0.849, F = 18.1(4, 28),$$

$$s = 0.458, \text{ AVRES} = 0.359, \text{ SDEP} = 0.568,$$

$$S_{PRESS} = 0.616, \text{ PRESS} = 10.6, \text{ Pres}_{av} = 0.459$$
 (6)

Table 3. Intercorrelation (|r|) among important descriptors

	pC_2	$pC_{2-}pC_1$	S_3	S_4	S_8	S_{12349}	S_{4589}	n	I_{OMe}	$I_{ m Ph/F}$	$I_{ m Furyl}$
pC_1	0.802	0.475	0.023	0.041	0.073	0.025	0.072	0.765	0.404	0.463	0.190
pC_2		0.144	0.049	0.149	0.221	0.055	0.196	0.357	0.474	0.219	0.435
$pC_{2-}pC_1$			0.110	0.152	0.206	0.123	0.171	0.744	0.030	0.444	0.326
S_3				0.941	0.775	0.995	0.857	0.082	0.475	0.070	0.032
S_4					0.937	0.950	0.976	0.061	0.427	0.169	0.011
S_8						0.807	0.989	0.067	0.353	0.228	0.045
S_{12349}							0.883	0.081	0.482	0.078	0.044
S_{4589}								0.046	0.391	0.214	0.034
n									0.269	0.418	0.179
I_{OMe}										0.115	0.132
$I_{ m Ph/F}$											0.205

Table 4. Results of leave-20%-out cross-validation^a

Eq. No.	Q^2	PRESS	
1	0.721	16.3	
2	0.720	16.3	
3	0.588	18.9	
4	0.576	19.5	
5	0.458	11.4	
6	0.446	11.7	

^a Compounds were deleted in five cycles in the following manner: 1, 6, 11,...,31; 2, 7, 12,..., 32; ...; 5, 10,...,30.

The predictor variables of Eqs. 5 and 6 are not much intercorrelated (Table 3). Eq. 6 could predict 49.6% and explain 68.1% of the variance of selectivity for binding with MT_2 over MT_1 . The variable S_{4589} in Eq. 6 indicates that electron density distribution of different atoms of the benzofuran ring is important for the selective binding with MT_2 over MT_1 receptor type. Moreover, the positive coefficient of n indicates the MT_2 selectivity increases on increase of number of CH_2 spacer (n) beside R_2 . Further, the positive coefficient of I_{OMe} indicates that the MT_2 selectivity increases in presence of methoxy group at R_1 . Again, the negative coefficient of I_{Furyl} denotes that MT_2 selectivity decreases if furyl substitution at R_3 position is present. The calculated and predicted selectivity values according to Eq. 6 are given in Table 1.

Leave-20%-out cross-validation was applied on all reported equations and the results are reported in Table 4.

The present QSAR study could throw some light on the physicochemical requirements of benzofuran derivatives for binding with MT₂ and MT₁ receptor types. The study also shows the utility of E-state index in developing statistically acceptable model having direct physicochemical significance. However, more data points covering wider features of substitution pattern need to be considered to reach a conclusion.

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References and notes

- Tan, D. X.; Reiter, R. J.; Manchester, L. C.; Yan, M. T.; El-Sawi, M.; Sainz, R. M.; Mayo, J. C.; Kohen, R.; Allegra, M.; Hardeland, R. Curr. Top. Med. Chem. 2002, 2, 181.
- 2. Reiter, R. J. Mol. Cell Endocrinol. 1991, 79, C153.

- Reiter, R. J.; Tan, D. X.; Mayo, J. C.; Sainz, R. M.; Leon, J.; Czarnocki, Z. Acta Biochim. Pol. 2003, 50, 1129.
- Tan, D. X.; Chen, L. D.; Poeggeler, B.; Manchester, L. C.; Reiter, R. J. Endocrine J. 1993, 1, 57.
- Cuzzocrea, S.; Reiter, R. J. Curr. Top. Med. Chem. 2002, 2, 153.
- 6. Cuzzocrea, S.; Zingarelli, B.; Gilard, E.; Hake, P.; Salzman, A. L.; Szabó, C. J. Pineal Res. 1997, 23, 106.
- Blask, D. E.; Sauer, L. A.; Dauchy, R. T. Curr. Top. Med. Chem. 2002, 2, 113.
- Masana, M. I.; Doolen, S.; Ersahin, C.; Al-Ghoul, W. M.; Duckles, S. P.; Dubocovich, M. L.; Krause, D. N. J. Pha. Exp. The. 2002, 302, 1295.
- Dubocovich, M. L.; Rivera-Bermudez, M. A.; Gerdin, M. J.; Masana, M. I. Front Biosci. 2003, 8, 1093.
- Spadoni, G.; Mor, M.; Tarzia, G. Biol. Signals. Recept. 1999, 8, 15.
- 11. Marot, C.; Chavatte, P.; Morin-Allory, L.; Viaud, M. C.; Guillaumet, G.; Renard, P.; Lesieur, D.; Michel, A. *J. Med. Chem.* **1998**, *41*, 4453.
- Rivara, S.; Mor, M.; Silva, C.; Zuliani, V.; Vacondio, F.; Spadoni, G.; Bedini, A.; Tarzia, G.; Lucini, V.; Pannacci, M.; Fraschini, F.; Plazzi, P. V. J. Med. Chem. 2003, 46, 1429.
- Spadoni, G.; Balsamini, C.; Diamantini, G.; Tontini, A.; Tarzia, G.; Mor, M.; Rivara, S.; Plazzi, P. V.; Nonno, R.; Lucini, V.; Pannacci, M.; Fraschini, F.; Stankov, B. M. J. Med. Chem. 2001, 44, 2900.
- Mor, M.; Spadoni, G.; Di Giacomo, B.; Diamantini, G.;
 Bedini, A.; Tarzia, G.; Plazzi, P. V.; Rivara, S.; Nonno,
 R.; Lucini, V.; Pannacci, M.; Fraschini, F.; Stankov, B. M.
 Bioorg. Med. Chem. 2001, 9, 1045.
- Wallez, V.; Durieux-Poissonnier, S.; Chavatte, P.; Boutin, J. A.; Audinot, V.; Nicolas, J. P.; Bennejean, C.; Delagrange, P.; Renard, P.; Lesieur, D. J. Med. Chem. 2002, 45, 2788.
- Kier, L. B.; Hall, L. H. Molecular Structure Description: The Electrotopological State; Academic: San Diego, 1999.
- 17. Hall, L. H.; Mohney, B.; Kier, L. B. Quant. Struct.-Act. Relat. 1993, 12, 44.
- Roy, K.; Pal, D. K.; Sengupta, C. Drug Des. Discov. 2001, 17, 207.
- Roy, K.; De, A. U.; Sengupta, C. Drug Des. Discov. 2002, 18, 33.
- Roy, K.; De, A. U.; Sengupta, C. *Indian J. Chem.* 1999, 38B, 942.
- 21. Roy, K.; Leonard, J. T. Bioorg. Med. Chem. 2004, 12, 745.
- Roy, K.; Chakraborty, S.; Saha, A. Bioorg. Med. Chem. Lett. 2003, 13, 3753.
- 23. CHEM DRAW ULTRA VERSION 5.0 is a program of ChambridgeSoft Corporation, USA.
- Ghose, A. K.; Crippen, G. M. J. Chem. Inf. Comput. Sci. 1987, 27, 21.
- 25. The GW-BASIC programs ELECTRO1, AUTOREG, RRR98, KRPRES1 and KRPRES2 were developed by Kunal Roy and standardized on known data sets.
- Snedecor, G. W.; Cochran, W. G. Statistical Methods;
 Oxford and IBH: New Delhi, 1967; pp 381–418.
- Wold, S.; Eriksson, L. In *Chemometric Methods in Molecular Design*; Waterbeemd, H. van de, Ed.; VCH: Weinheim, 1995; pp 312–317.